CLINICAL AND HEMATOLOGICAL ASPECTS OF DOGS NATURALLY INFECTED WITH Ehrlichia spp. AND Leptospira interrogans

ASPECTOS CLÍNICOS E HEMATOLÓGICOS DE CÃES NATURALMENTE INFECTADOS COM Ehrlichia spp. E Leptospira interrogans

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ABSTRACT: Ehrlichiosis and leptospirosis are frequent diseases in tropical countries, although they are not always considered as differential diagnosis. The purpose of this work was to evaluate clinical signs and hematological alterations in dogs reactive to leptospirosis, among them those that presented morulae of Ehrlichia spp. in peripheral blood, and to recognize the frequency of Leptospira serovars by means of the microscopic agglutination test. Blood samples were used from 50 dogs presenting morulae of Ehrlichia spp. in blood extensions collected from the ear marginal capillaries. The clinical signs frequently observed were apathy (100.00%), anorexia/dysorexia (100.00%), adypsia (83.33%), ocular secretion (66.66%), reactive lymph nodes (66.66%), diarrhea (58.33%), vomiting, anemia and fever (41.66%). Albeit unspecific, anemia and thrombocytopenia are frequent hematological findings in canine ehrlichiosis and are present in both diseases. Of the 50 dogs with ehrlichiosis, 12 (24.00%) were reactive to leptospirosis, and the most frequent serovars were Pomona (33.33%), followed by Canicola (27.77%) and Icterohaemorrhagiae (22.22%). The results suggest the need to establish a differential diagnosis between these diseases that may occur concomitantly.


INTRODUCTION

The canine population’s growth has been favored by urbanization and by social changes in the population (SOUZA et al., 2002). Hence, dogs participate effectively in the epidemiological chain of a variety of zoonosis since they are pets that have close contacts with humans (BRASIL, 2002).

Ehrlichiosis, which is considered a zoonosis, is caused by an obligate intracellular parasite. Several species of ehrlichia can contaminate dogs, including Ehrlichia canis – the agent of canine monocytic ehrlichiosis, Ehrlichia chaffeensis – the agent of human monocytic ehrlichiosis, and Ehrlichia ewingii – the agent of canine granulocytic ehrlichiosis (DAGNONE et al., 2001; DUMLER et al., 2001).

In Brazil, E. canis is imputed to be responsible for the most pathogenic canine monocytic ehrlichiosis frequently associated with the clinical disease (LAPPIN, 2010). The disease is considered endemic, mainly in urban areas, where there is an abundant occurrence of populations of the vector tick, Rhipicephalus sanguineus (LABRUNA; PEREIRA, 2001). In the state of Minas Gerais, the disease was reported in dogs for the first time in the city of Belo Horizonte (COSTA et al., 1973).

The diagnosis can be made based on the detection of intracellular clusters of the microorganism contained within a membrane, called morulae, which are present in the leukocytes in peripheral blood smears during the acute phase (STILES, 2000) or by means of serology using Indirect Immunofluorescence, ELISA or PCR techniques, and by isolation and culture (ANDEREG; PASSOS, 1999). Serology and nested PCR are suitable assays for the confirmation of canine ehrlichiosis, but they should complement clinical and hematological exams (CONH, 2003; NAKAGHI et al., 2008). Therefore, serology is important in the phases in which there is no parasitemia, and also allows for the identification of the species involved.

Visualization of morulae in peripheral blood provides a definitive diagnosis, but it is a slow process that also yields many false-negative results. Although it is corroborative, the hematozoon examination is severely limited by its low sensitivity, since it may be difficult to observe corpuscles and morulae in the subacute and chronic phases due to low parasitemia, thus making it...
advisable to use other diagnostic methods (MOREIRA et al., 2005).

Another important zoonosis is leptospirosis whose easy transmission leads to high morbidity according to public health warnings. Among domestic animals in urban areas, dogs are the main source of infection for humans because of their direct contact and the fact that they can asymptomatically eliminate live leptospirae in their urine for several months (MAGALHÃES et al. 2006; ADLER; MOCTEZUMA, 2010).

The diagnosis is based on clinical and epidemiological information, confirmed by laboratorial exams, and the microscopic agglutination test with live antigens is recommended by the World Health Organization (WHO, 2003).

Routine exams at the Federal University of Uberlândia Veterinary Teaching Hospital of the College of Veterinary Medicine show frequent diagnosis of animals with ehrlichiosis. However, the clinical diagnosis confirmed by the detection of morulae in peripheral blood smears interrupts the search for other etiological agents that may be present in the clinical picture under evaluation.

The lack of a differential diagnosis due to similar clinical factors leads to inadequate treatment, since it does not eliminate the renal carrier stage of *Leptospira* spp. In view of this concern and the possibility of other diseases affecting an animal at the same time, it was decided to verify the occurrence of animals reactive to leptospirosis among those testing positive for ehrlichiosis.

The aim was to evaluate clinical signs and hematological abnormalities in dogs reactive to *Leptospira* spp. among those that presented *Ehrlichia* spp. morulae in peripheral blood. Another aim was to ascertain the frequency of *Leptospira* serovars by means of the Microscopic Agglutination Test (MAT).

**MATERIAL AND METHODS**

The study involved 50 dogs of both sexes, of various breeds and ages, treated at the Veterinary Teaching Hospital of the Federal University of Uberlândia, MG, Brazil from November 2007 to April 2008. These animals, which had not previously received any kind of treatment, presented morulae of *Ehrlichia* spp. in blood samples drawn from the ear marginal capillaries. The blood smears were stained by the May-Grünwald Giemsa (FELDMAN et al., 2000) method and were used for a study of hemoparasites and a differential leukocyte count.

The clinical signs of all the dogs were evaluated according to the animal’s clinical chart tracked by the number of its medical record.

After antisepsis with alcohol iodine, 5 mL of blood were collected from the accessory cephalic or external jugular vein, using 5 mL syringes and disposable 25x7 needles for the hemogram. Two mL of blood were then transferred to a silicone coated vacutainer tube containing anticoagulant (0.1 mL of 10% EDTA – ethylene-diamine-tetra-acetic acid), and 3 mL were placed in a tube without anticoagulant for serum extraction.

The hemograms were performed at the Laboratory of Clinical Pathology of the Veterinary Teaching Hospital of FAMEV, UFU, by the electronic method, using an ABC VET® animal blood counter. The tubes containing blood samples without anticoagulant were sent to the Laboratory of Contagious Infectious Diseases of the same institution for the leptospirosis serum screening test. The samples were centrifuged at 2,500 rpm for 5 min. The serum was then extracted with a micropipette and transferred to conical-bottom polypropylene tubes, which were stored at -20°C until the moment of the exam.

The diagnostic method adopted for the serology of leptospirosis was MAT, considered the golden standard, which detects serum antibodies starting seven days after infection (BRASIL, 1995; LABRUNA; PEREIRA, 2001). The serovars tested were Hardjo, Canicola, Pomona, Wolffii, Icterohaemorrhagiae, Grippotyphosa and Pyrogenes. Antigens were prepared from matrices transplanted weekly in a STUART (Difco®) culture medium enriched with 10% rabbit serum, stored in an incubator at 30°C and used close to the sixth day of incubation.

The coefficient of prevalence of the animals testing positive for leptospirosis was determined, as was the most frequent serovar and the clinical signs with the highest occurrence (THRUFSFIELD, 2004). The results were analyzed by descriptive statistics, using Student’s parametric test for two independent samples (t test) with a 5% level of significance, for the comparison of two means according to Ayres et al. (2007).

The research was developed according to the standards established by the institution’s ethics committee (015/08).

**RESULTS**

Among the 50 samples containing morulae of *Ehrlichia* spp. in their peripheral smears, 12 were
reactive to leptospirosis in the MAT test, resulting in a prevalence of 24.00%.

The frequencies of serovars showed 33.33% of dogs reactive to the serovar Pomona, 27.77% positive for Canicola, 22.22% for Icterohaemorrhagiae, 11.11% for Wolffi and 5.55% for Hardjo. No animals were found to be serum reactive to the serovars Pyrogenes and Grippotyphosa. Coagglutination occurred when the same dog was reactive to more than one serovar.

The main hematological alterations of the 50 dogs with ehrlichiosis were anemia and thrombocytopenia, as Table 1.

Table 1. Mean, standard deviation, and amplitude of variation of the hematological parameters of 50 domestic dogs naturally infected with *Ehrlichia* spp. and treated at the Veterinary Teaching Hospital of the Federal University of Uberlândia, MG, Brazil, 2008.

<table>
<thead>
<tr>
<th>Parameters evaluated</th>
<th>Mean</th>
<th>Standard deviation</th>
<th>Minimum</th>
<th>Maximum</th>
<th>Reference values*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hematocrit %</td>
<td>29.04</td>
<td>11.23</td>
<td>7.70</td>
<td>57.20</td>
<td>37-55</td>
</tr>
<tr>
<td>Erythrocytes x 10^6/µL</td>
<td>4.52</td>
<td>1.71</td>
<td>1.04</td>
<td>8.25</td>
<td>5.5-8.5</td>
</tr>
<tr>
<td>Hemoglobin g/dL</td>
<td>9.45</td>
<td>3.77</td>
<td>2.50</td>
<td>20.00</td>
<td>12-18</td>
</tr>
<tr>
<td>Platelets /µL</td>
<td>108,222</td>
<td>96,795</td>
<td>14,000</td>
<td>443,000</td>
<td>200,000-500,000</td>
</tr>
<tr>
<td>Leukocytes /µL</td>
<td>9,008</td>
<td>6,484</td>
<td>1,500</td>
<td>41,200</td>
<td>6,000-17,000</td>
</tr>
<tr>
<td>Segmented neutrophils /µL</td>
<td>4,971</td>
<td>3,923</td>
<td>285</td>
<td>24,720</td>
<td>3,000-11,500</td>
</tr>
<tr>
<td>Lymphocytes /µL</td>
<td>2,168</td>
<td>10,604</td>
<td>261</td>
<td>10,604</td>
<td>1,000-4,800</td>
</tr>
<tr>
<td>Monocytes /µL</td>
<td>739</td>
<td>883</td>
<td>112</td>
<td>5,061</td>
<td>150-1,350</td>
</tr>
<tr>
<td>Basophils /µL</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>Rare</td>
</tr>
</tbody>
</table>

* Meinkoth; Clinkenbeard (2000).

In this study, it was possible to compare the mean hematological values of the dogs infected only with *Ehrlichia* spp. against those infected with both *Ehrlichia* spp. and *Leptospira interrogans* (Table 2). Figure 1 describes the complaints and clinical signs found in the dogs.

Table 2. Comparison of the mean hematological values of dogs naturally infected only with *Ehrlichia* spp. and those infected with *Ehrlichia* spp. and *Leptospira interrogans*.

<table>
<thead>
<tr>
<th>Parameters evaluated</th>
<th>Mean values</th>
<th>Reference values*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td><em>Ehrlichia</em> spp.</td>
<td><em>Ehrlichia</em> spp.+L. interrogans</td>
</tr>
<tr>
<td>Hematocrit %</td>
<td>30.39a</td>
<td>24.82b</td>
</tr>
<tr>
<td>Erythrocytes x 10^6/µL</td>
<td>4.73a</td>
<td>3.87b</td>
</tr>
<tr>
<td>Hemoglobin g/dL</td>
<td>9.88a</td>
<td>8.07b</td>
</tr>
<tr>
<td>Platelets /µL</td>
<td>102,107a</td>
<td>127,583b</td>
</tr>
<tr>
<td>Leukocytes /µL</td>
<td>8,434a</td>
<td>10,825b</td>
</tr>
<tr>
<td>Segmented neutrophils /µL</td>
<td>4,620a</td>
<td>6,083b</td>
</tr>
<tr>
<td>Lymphocytes /µL</td>
<td>2,037a</td>
<td>2,580a</td>
</tr>
<tr>
<td>Monocytes /µL</td>
<td>663a</td>
<td>981a</td>
</tr>
<tr>
<td>Basophils /µL</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

*Meinkoth; Clinkenbeard (2000); (a; b) different letters on the same line indicate statistically different means (p<0.05).
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Figure 1. Clinical signs presented by dogs infected naturally with *Ehrlichia* spp. and *Leptospira interrogans*

**DISCUSSION**

Ehrlichiosis and leptospirosis can occur simultaneously in the same host without being properly diagnosed, because ehrlichiosis is an opportunistic agent that develops when the immunological system is imbalanced. In addition to being immunosuppressant, ehrlichiosis can trigger medullary aplasia and drastically reduce the defense cells, thus allowing the installation of other pathogens such as *Leptospira interrogans*.

Of the 50 dogs evaluated in this study, 12 were MAT reactive. The prevalence of leptospirosis (24.00%) should not be compared with that found by other researchers because no similar evaluation was found in the literature, since the methodologies employed were different.

The frequencies of serovars found were differed from those reported by other authors (FURTADO et al., 1997; ÁVILA et al., 1998; ALVES et al., 2000), who found serum prevalence for Autumnalis, Butembo, Grippotyphosa, Australis, Canicola and Icterohaemorrhagiae. Infecting serovars depend of the assessed species, the contactant animals, the region evaluated, the presence of hosts, and maintenance of the agent in the environment, and there may be differences of serum prevalence in evaluated locations (FURTADO et al., 1997).

The high occurrence of the serovar Canicola confirms that dogs are an important source of contamination for humans because they are the main host of this serovar, which adapts easily to the canine renal tissue and can be eliminated for long periods after infection. The discovery of serovars not commonly observed in current commercial vaccines is cause for concern, since there is no cross-immunity between serovars.

The most commonly serovars in canine leptospirosis have been Icterohaemorrhagiae and Canicola, but others serovars have emerged as significant cause of the disease. An example is the serovars Grippotyphosa and Pomona, whose prevalence in dogs has increased and which are important infectious causes of acute renal failure (ETTINGER; FELDMAN, 2004). The serovar Pomona, which was the most prevalent in this study, has been reported by other authors at only 1.6% and 0.74% (PETRUCHI et al., 2002). However, another research (TESSEROLLI et al., 2008) found a prevalence of 32.27% (193/598) with serovars of higher prevalence reported as Copenhageni (71.50%) and Canicola (6.74%), accompanied by Icterohaemorrhagiae (2.08%).
The clinical and hematological aspects of dogs infected with *E. canis* described Nakaghi et al. (2008) were apathy (60.70%), anorexia (56.70%), pale mucosae (43.30%), fever (43.30%), lymphadenomegaly, hepatoomegaly and/or splenomegaly (43.30%), and uveitis (40.00%), followed by petechial hemorrhages or suffusions (33.30%), as the main findings in 30 dogs clinically diagnosed with ehrlichiosis.

During the acute phase of canine monocytic ehrlichiosis, Rungsipipat et al. (2009) noted fever, depression, anorexia, lymphadenopathy, diffuse bleeding, episcleritis and conjunctivitis, clinical signs which were also observed in this study.

The immune response that triggers the host to injury before the agent generates an increase in the production of interleukin-1 and antigen expression in immune cells, mainly in B lymphocytes. There is also a production of exogenous pyrogens rickttsia resulting in hyperthermia and break the balance of systems.

Shipov et al. (2008) established clinical signs of prognosis in canine ehrlichiosis and as observed in this study were pale mucous membranes, weakness and spontaneous bleeding. These signs were related mainly to the group of dogs that survived and had a worse prognosis, constituting a risk factor for mortality in ehrlichiosis in dogs. De Castro et al. (2004) described yet ascitis and splenomegaly.

The clinical signs frequently associated with canine leptoospriosis depend on several factors such as the virulence of the serovar and the host's immunological status and age. The mildest forms evolve to fever, anorexia, vomiting, dehydration and apathy and the chronic form may be unapparent, culminating in chronic renal insufficiency (SANTIM et al., 2006). The clinical changes in the chronic phase might occur because of the reactions against the agent, like as the narrow hypoplasia/aplasia. These abnormalities may be explained due to narrow suppression that occurs with depletion of cell lines (LAPPIN, 2010). Geisen et al. (2007) had determined that the clinical signals commonly associated with the infection by *Leptospira interrogans* in the dogs had been anorexia, vomiting, lethargy and weakness.

According to the hemogram (Table 1), the mean values of leukocytes, segmented neutrophils, lymphocytes and monocytes found both in dogs positive for ehrlichiosis and leptoospriosis and in those positive only for ehrlichiosis were within the normal parameters. The means obtained from the erythrocyte count, measurement of the concentration of hemoglobin and the hematocrit values were lower than the normal values (FELDMAN et al., 2000; MEINKOTH; CLINKENBEARD, 2000) characterized by anemia normoregenerative normocytic and normochromic.

Thrombocytopenia, nonregenerative anemia, eosinopenia, and nuclear deviation of neutrophils to the left are frequent hematological alterations in canine ehrlichiosis (MENDONÇA et al., 2005), which were also observed in this study. In a clinical study involving 30 dogs with ehrlichiosis, 28 were confirmed through laboratory exams, the most frequent pathological and clinical findings being anemia, leukopenia and thrombocytopenia (SHIPOV et al., 2008; TESSEROLLI et al., 2008).

The results of this research agree with De Castro et al. (2004) and Rungsipipat et al. (2009) who described normoregenerative normocytic anemia, thrombocytopenia and leukopenia in blood counts in dogs with monocytic ehrlichiosis. The thrombocytopenia can be explained by the immune-mediated destruction, sequestration or by decreased production, vasculitis and platelet function abnormalities (LAPPIN, 2010).

The hematological alterations commonly observed in leptoospriosis are intense leukocytosis, neutrophilia and varied degrees of anemia. Leukopenia can be established in the initial phase of the disease during leptoospriemia and evolve into leukocytosis with left shift as the disease progresses. Severely affected dogs may present thrombocytopenia (GREENE et al., 2006).

Leukopenia is common in canine ehrlichiosis, but in the acute phase of the disease it may occur leucopenia, followed by leukocytosis with neutrophilia and monocytosis (LAPPIN, 2010). In this phase, dogs with ehrlichiosis may present...
hematological results similar to those of dogs with leptospirosis, since both may present leukocytosis, thrombocytopenia and anemia. However, the occurrence of these alterations depends on the host’s immunological status.

A clinical improvement can be achieved in cases of leptospirosis that were not correctly diagnosed, since the drug of choice for the treatment of ehrlichiosis is doxycycline, which is also recommended for the treatment of leptospirosis, although it does not eliminate the renal carrier status, but decrease the excretion of leptospires in urine. Therefore, doxycycline can treat the two diseases concomitantly and lead to the remission of the symptoms (ETTINGER, FELDMAN, 2004). Clinicians should therefore be aware, since *Leptospira interrogans* may remain in the organism of the infected animal for a long time, disseminating the agent in the environment and exposing other animals, including humans, to the disease.

Albeit unspecific, the complaints and clinical signs of apathy, anorexia/dysorexia, adipsia, ocular secretion, lymphadenomegaly, diarrhea, vomiting and hyperthermia are frequently found in canine ehrlichiosis and are present in dogs with leptospirosis and ehrlichiosis, as well as the hematological alterations of anemia and thrombocytopenia. This study detected the occurrence of anti-*Leptospira* agglutinins in dogs infected naturally by *Ehrlichia* spp. It is suggested that, when dogs exhibit clinical and hematological aspects of ehrlichiosis, a differential diagnosis should be done for leptospirosis, since they may occur concomitantly, aggravating the possibility of zoonosis.

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REFERENCES


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